

Total synthesis of beauveriolide I

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Abstract—The first total synthesis of beauveriolide I (**1a**), a selective ACAT inhibitor, is described. The key steps in this synthesis involved a diastereoselective aldol condensation sequence and a macrocyclization.

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Beauveriolide I (**1a**) was first isolated in 1975 from the mycelium of the strain *Beauveria* sp.¹ Its structure was elucidated as *cyclo*-[(3*S*,4*S*)-3-hydroxy-4-methyl-octanoyl-L-phenylalanyl]-D-leu-Cine, a member of the cyclodepsipeptide family,² by spectral analyses and chemical degradation.

Recent studies show beauveriolide I (**1a**) and III (**1b**) can cause a reduction in both number and size of cytosolic lipid droplets in macrophages without any cytotoxic effect on macrophages.^{3,4} Among these beauveriolides, beauveriolide I (**1a**) and III (**1b**) are the most potent inhibitors of lipid droplets formation in mouse macrophages. They inhibit Acyl-CoA: cholesterol acyltransfer-

ase (ACAT) activity specifically, resulting in the blockage of cholesterol ester (CE) synthesis, leading to a reduction of lipid droplets in macrophages.⁵

Intrigued by beauveriolides' bioactivity and interested in their SAR studies, we began the synthetic study and achieved the first total synthesis of beauveriolide I (**1a**). The retrosynthetic analysis is shown in Figure 1.

As the macrocyclization can be achieved by either amide bond or ester bond formation, the major endeavor is the synthesis of the fragment **2** [(3*S*,4*S*)-3-hydroxy-4-methyloctanic acid], which is a common intermediate in beauveriolide I (**1a**), III (**1b**) and other beauveriolides.⁶

Diastereoselective aldol condensation has been utilized for the synthesis of compound (**2**). Reaction of 3-benzyl-oxypropionaldehyde⁷ and chiral imide (**5**)⁸ under Crimmins' condition⁹ furnished *syn* aldol (**6a**)¹⁰ in excellent yield and diastereoselectivity. Installation of TBDMS protecting group followed by reduction with LiBH₄ gave alcohol (**7**)¹¹. Alcohol (**7**) was oxidized under Swern

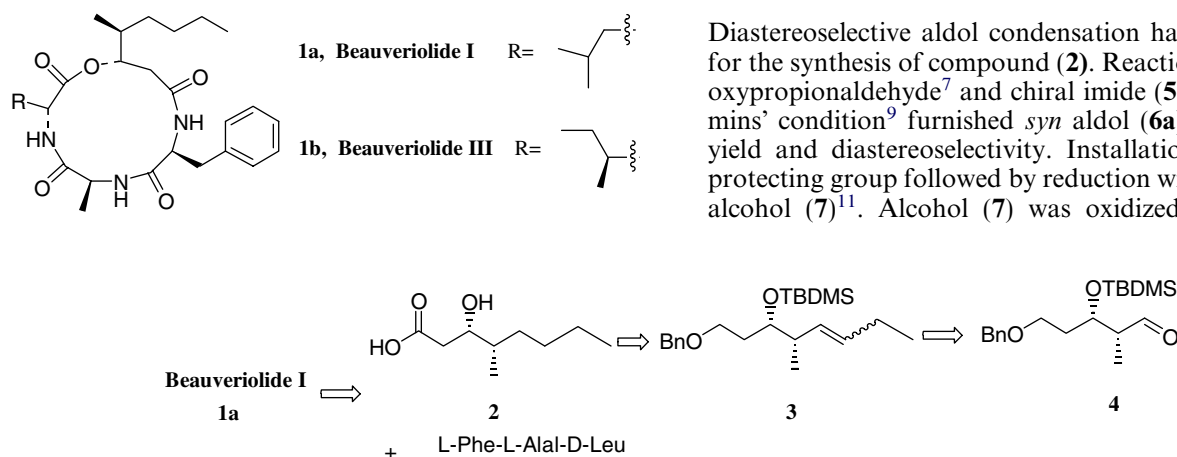
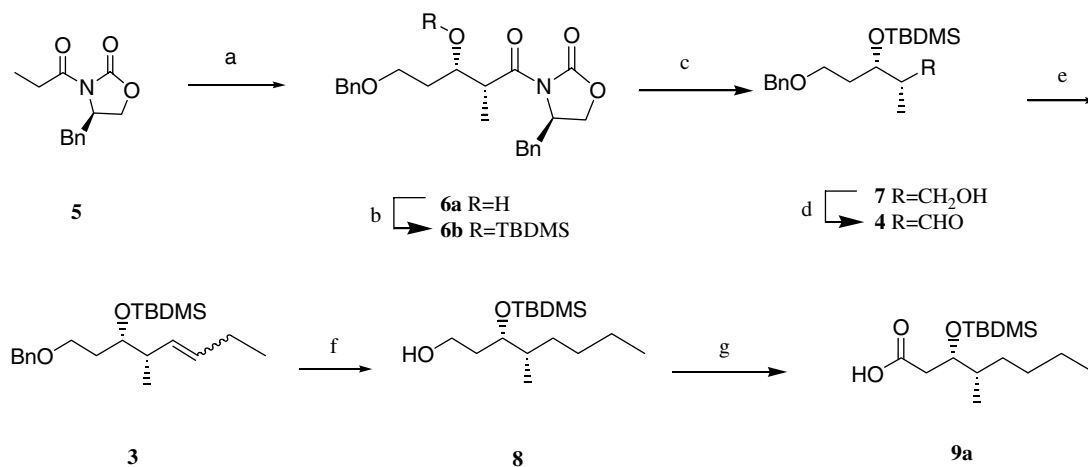


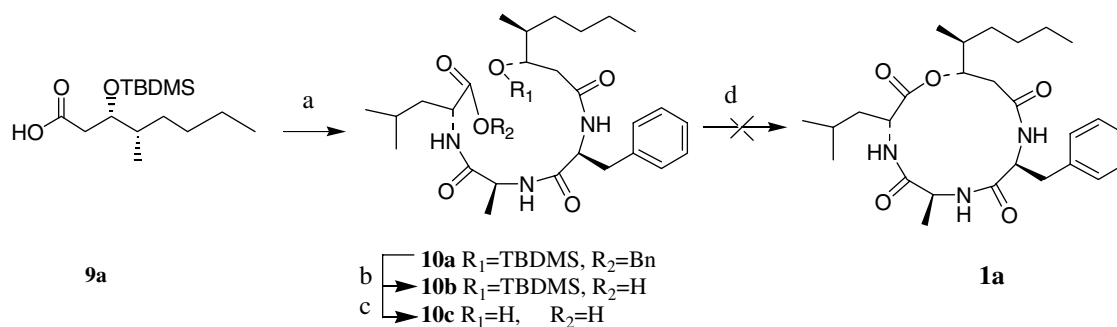
Figure 1. Retrosynthetic analysis of beauveriolide I.

Keywords: ACAT inhibitor; Beauveriolide I; Total synthesis.

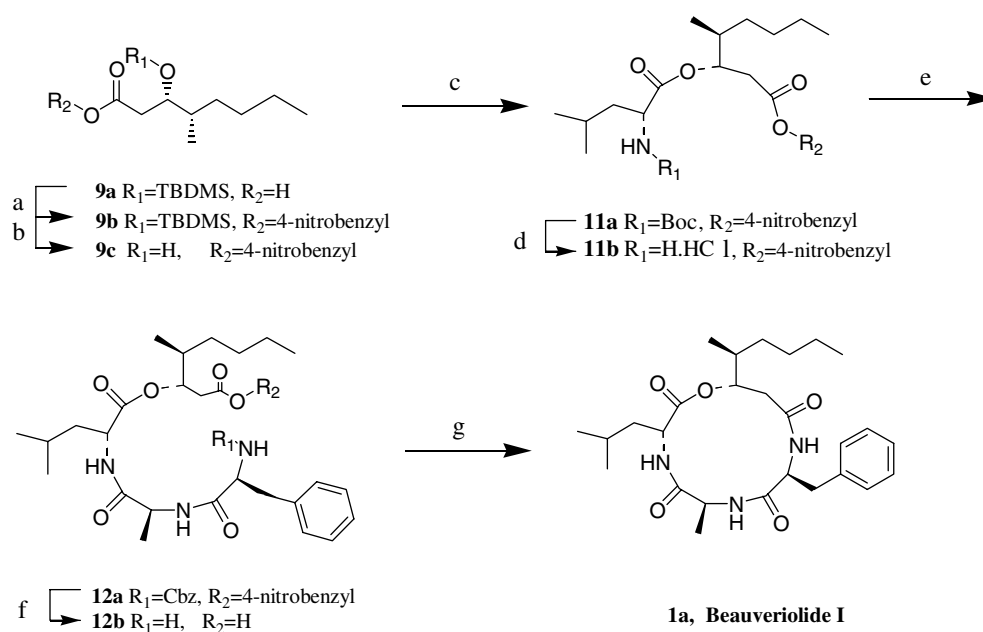
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Scheme 1. Reagents and conditions: (a) 3-benzoyloxypropionaldehyde, TiCl₄, DIPEA, NMP, CH₂Cl₂, −5 °C to rt, 3.5 h, 97%; (b) TBDMSCl, imidazole, DMF, rt, 30 h, 79%; (c) 2 M LiBH₄, H₂O, Et₂O, 0 °C to rt, 73%; (d) 1. (COCl)₂, DMSO, CH₂Cl₂, −78 °C, 2. NEt₃, −78 °C to rt, 90%; (e) [Ph₃PCH₂CH₂CH₃]Br, 1.6 M *n*-BuLi, THF, rt, 2 h, 91%; (f) H₂, 10% Pd–C, EtOH, rt, 82%; (g) NaIO₄, RuCl₃(cat.), CCl₄/CH₃CN/H₂O (v/v/v = 2/2/3), rt, 2 h, 78%.



Scheme 2. Reagents and conditions: (a) L-Phe-L-Ala-D-Leu-OBn, DCC, CH₂Cl₂, 0 °C to rt, 6 h; (b) H₂, 10% Pd–C, THF; (c) HOAc, heating; (d) 2-methyl-6-nitrobenzoic anhydride, DMAP, toluene.



Scheme 3. Reagents and conditions: (a) 4-nitrobenzyl bromide, DIPEA, DMF, rt, 7 h, 66%; (b) 65–75% HF–Py, THF, rt, 20 h, 70%; (c) DCC, DMAP(cat.), CH₂Cl₂, rt; (d) HCl/EtOAc, 4 h, 0 °C to rt; (e) *N*-Cbz-L-Phe-L-Ala, EDCI, DIPEA, HOSu(cat.), DMF, 0 °C to rt; (f) H₂, 10% Pd–C, THF/isopropanol (v/v = 1/2.5), rt; (g) BOP, DMAP, CH₃CN, rt, 68%.

condition,¹² and the resulting aldehyde (**4**) was allowed to react with propyltriphenylphosphonium bromide in the presence of *n*-BuLi to afford the corresponding olefinic isomers (**3**). Hydrogenation of the olefinic bond and removal of the benzyl protecting group were accomplished in one step over a catalytic amount of 10% Pd/C. Oxidation with NaIO₄ in the presence of RuCl₃¹³ gave the desired acid (**9a**) in 89% yield (Scheme 1).

With the acid (**9a**) in hand, we tried to perform the macrocyclization by ester bond formation as shown in Scheme 2, where the suitably protected liner precursor **10a** was synthesized by coupling **9a** with L-Phe-L-Ala-D-Leu-OBn in the presence of DCC and DMPA, followed by hydrolysis to remove the TBDMS group. However, unfortunately, the desired compound **1a** could not be obtained under various conditions.¹⁴

We therefore selected the macrocyclization through amide bond formation. The synthetic sequence of compound **12a** was shown in Scheme 3. Protection of **9a** using 4-nitrobenzyl bromide in the presence of DIPEA in DMF yielded ester **9b**. Removal of the TBDMS protecting group by treating with HF-pyridine afforded compound **9c**,¹⁵ which was coupled with *N*-Boc-D-Leu¹⁶ by using DCC and catalytic amount of DMAP to give diester (**11a**). After removing the Boc group the corresponding protected tetradepsipeptide (**12a**) could be obtained by coupling with the *N*-Cbz-L-Phe-L-Ala¹⁷ by using EDCI in the catalysis of HOSu. Subsequent reductive removal of the 4-nitrobenzyl group and the Cbz group gave the deprotected tetradepsipeptide (**12b**). The macrocyclization was performed successfully using BOP in acetonitrile under highly dilute conditions (5.3×10^{-3} mol/L)¹⁸ to furnish beauveriolide I (**1a**)¹⁹ in 68% yield. The structure of the product (**1a**) was confirmed by NMR and mass analysis for its structural data and rotation value matching well with the natural product.^{2,20}

In conclusion, the first total synthesis of beauveriolide I (**1a**), an antibiotic from the culture broth of fungal *Beauveria* sp. FO-6979 was achieved. The synthesis is flexible and amenable to other analogues for SAR studies.

References and notes

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- Analytical data of compound **6a**: pale-yellow oil, $[\alpha]_D^{25}$ –44.3 (*c* 0.3, CHCl₃). IR (film): 1778, 1695, 1387, 1209, 1109 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.24–1.28 (m, 2H), 1.29 (d, 3H, *J* = 4.5 Hz), 1.71–1.79 (m, 1H), 1.83–1.93 (m, 1H), 2.77 (dd, 1H, *J* = 13.2, 9.6 Hz), 3.25 (dd, 1H, *J* = 13.2, 3.3 Hz), 3.62–3.70 (m, 2H), 3.78–3.86 (m, 1H), 4.09–4.22 (complex, 3H), 4.52 (s, 2H), 4.64–4.72 (m, 1H), 7.19–7.38 (complex, 10H). FABMS: 398 (M+H⁺), HR-FABMS: calcd for C₂₃H₂₈NO₅: (M+H⁺), 398.1967, found: 398.1979.
- Analytical data of compound **7**: $[\alpha]_D^{25}$ –8.5 (*c* 1.3, CHCl₃). ¹H NMR (500 MHz, CDCl₃): δ 0.054 (s, 3H), 0.096 (s, 3H), 0.81 (d, 3H, *J* = 7 Hz), 0.88 (s, 9H), 1.78–1.84 (m, 2H), 1.98 (br, 1H), 3.50–3.56 (m, 3H), 3.69 (t, 1H, *J* = 9.5 Hz), 3.71–3.96 (m, 1H), 4.49 (q, 2H, *J* = 11.5 Hz), 7.26–7.36 (complex, 5H). ¹³C NMR (125 MHz, CDCl₃) δ –4.8, –4.5, 12.4, 17.9, 25.8, 32.2, 39.9, 65.8, 67.1, 72.8, 73.0, 127.6, 127.7, 128.4, 138.4.
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- Analytical data of compound **9c**: pale-yellow oil, ¹H NMR (400 MHz, CDCl₃): δ 0.89 (t, 3H, *J* = 6.4 Hz), 0.92 (d, 3H, *J* = 6.4 Hz), 1.11–1.54 (m, 7H), 2.13 (br, 1H), 2.51–2.57 (complex, 2H), 3.98 (dt, 1H, *J* = 4.4, 8.4 Hz), 5.25 (s, 2H), 7.52 (d, 2H, *J* = 8.4 Hz), 8.23 (d, 2H, *J* = 8.4 Hz). ¹³C NMR (100 MHz, CDCl₃) δ 14.0, 14.2, 22.9, 29.4, 32.4, 38.1, 38.8, 64.9, 71.3, 123.8, 128.4, 142.9, 147.7, 172.8. FABMS: 310.2 (M+H⁺), HR-FABMS: calcd for C₁₆H₂₄NO₅: (M+H⁺), 310.1654, found: 310.1664.
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- Analytical data of compound **1a**: colorless needles, mp 242–244 °C (from MeOH); $[\alpha]_D^{25}$ –23.7 [*c* 0.7, CHCl₃–MeOH (4:1)]; IR (KBr): 3309, 1726, 1684, 1643, 1537 cm⁻¹. ¹H NMR (500 MHz, CDCl₃–CD₃OD = 4:1): δ 0.79 (t, 3H, *J* = 6.5 Hz), 0.80 (d, 3H, *J* = 6.5 Hz), 0.83 (d, 3H, *J* = 6.0 Hz), 0.85 (d, 3H, *J* = 5.5 Hz), 0.93–1.00 (m, 1H), 1.07–1.15 (m, 1H), 1.18 (d, 3H, *J* = 7.0 Hz, overlapped with 2H signal), 1.20–1.28 (m, 1H), 1.30–1.36 (m, 1H), 1.41–1.50 (m, 3H), 2.01–2.04 (m, 1H), 2.36 (dd, 1H, *J* = 14.0, 9.5 Hz), 2.42 (dd, 1H, *J* = 14.0, 5.5 Hz), 2.90 (dd, 1H, *J* = 13.5, 8.0 Hz), 2.99 (dd, 1H, *J* = 13.5, 8.5 Hz), 3.81 (dd, 1H, *J* = 14.0, 7.0 Hz), 4.16 (t, 1H, *J* = 8.0 Hz), 4.53–4.56 (m, 1H), 4.84–4.88 (m, 1H), 6.87–7.24 (m, 5H). ¹³C NMR (125 MHz, CDCl₃–CD₃OD = 4:1) δ 13.4, 14.5, 15.0, 21.7, 22.5, 24.5, 29.0, 30.3, 35.1, 35.3, 35.5, 40.8, 49.0, 52.3, 56.6, 76.0, 126.5, 128.1, 128.6, 136.0, 169.4, 171.0, 171.3, 171.8. FABMS: 488.3 (M+H⁺), HR-FABMS: calcd for C₂₇H₄₂N₃O₅: (M+H⁺), 488.3124, found 488.3147.
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